

Faculty of Science & Technology

Choice Based Credit System

Subject: BIOCHEMISTRY

(Additional Interdisciplinary Course)

Name of the Course: B. Sc. Part-II (Sem. III & IV)

(Syllabus to be implemented from June 2023)

SYLLABUS FOR B. Sc – II (BIOCHEMISTRY) CHOICE BASED CREDIT SYSTEM (CBCS)

Structure of the Course:

- Structure of B.Sc. course in faculty of science has total of 06 semesters for 3 years.
- B.Sc.-II comprises of total two semesters. Each semester will have **two** theory papers of 80 marks for university external examination and 20 marks for internal examination for eachpaper.
- At the end of academic year i.e. semester IV the practical examination will beconducted. The evaluation of practical exam is of 80 marks for university external practical examination and 20 marks for internal practical examination.
- The titles and marks distribution for each paper are asunder.

Semester	Paper No.	Title of Paper	Total Lectures	Ex	Examination		Total Credit
				Univ. Exam	Internal Exam	Total Marks	
Semester II	II	Biomolecules	30	40	10	50	02
	II	Biochemical Technique	s 30	40	10	50	02
Semester IV	/ III	Nutrition and Metabolism	30	40	10	50	02
	IV	Molecular Biochemistry and Diseases	30	40	10	50	02
Practical	Ι	Biochemistry		160	40	200	04

• University Examination

Theory Paper I : 40Marks
 Theory Paper II : 40Marks
 Theory Paper III : 40Marks
 Theory Paper IV : 40Marks
 Fractical : 160Marks

Practical paper has 160 marks for external university practical examination. Duration of practical examination is **TWO days**. Out of 160 marks for external university practical examination, the mark distribution is as follows.

Practical Course

Part I (80 Marks)

1)	Chromatographic experiments	20
2)	Colorimetric experiments	20
3)	Preparation experiments	20
4)	Study of an Enzyme	20

(activity quantification/effect or temperature/pH/substrate concentration).

	I	Part II	(80 Marks)
1)	Volumetric Estimations		20
2)	Qualitative Analysis		20
	a) Carbohydrates/Lipids		
	b) Enzymes/ Urine (Abnorm	al components)	
3)	Journal		20
4)	Study Tour		10
5)	Oral		10

• Continuous Internal Assessment:

- 1) Each theory paper has 10 marks for internal examination. There will be 05 marks for unit test and 05 marks for home assignment.
- 2) Practical paper has 40 marks for internal examination.

SEMESTER-III PAPER - I: BIOMOLECULES

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- Students learn about the components and structure of carbohydrate molecules.
- Students learn about the amino acids and their general classification.
- Students learn about the components and structure of protein.
- Students learn about the components and general classification of enzymes.
- Students learn about the components and structure of lipids.
- Students learn about the components and structure of nucleic acids.
- Students learn about the types and properties of vitamins.

SEMESTER-III PAPER - I: BIOMOLECULES

Total m	arks: 50 (40+10) Credits: 2 Contact ho	urs: 30
Unit	Content	Lecture
No.		S
Unit I	 CARBOHYDRATES: (06 L) Definition, classification, structures of- A) Monosaccharides- aldoses & ketoses i) Trioses- glyceraldehyde & dihydroxy acetone ii) Tetroses-erythrose, erythrulose iii) Pentoses-ribose, ribulose, xylose, xyluose iv) Hexoses-glucose, mannose, galactose, fructose. Reactions of monosaccharides, Derivatives of monosaccharides B) Oligosaccharides: glycoside bond, maltose, isomaltose, sucrose, cellobiose C) Polysaccharides- starch, glycogen, cellulose AMINO ACIDS: (04 L) Classification of amino acids. Properties of amino acids: physical properties, chemical properties, zwitterions, pI, ninhydrin reaction. PROTEINS & ENZYMES: (05 L) Formation of peptide bond, definition of proteins. i) Types of protein: Simple proteins (albumin & globulin), Complex proteins & Derived proteins ii) Structure of proteins: Primary, Secondary (α helix & β pleated), Tertiary & Quaternary structure, Forces involved in stabilizing native structure of protein. Enzymes - Definition, apoenzyme, coenzyme, holoenzyme, prosthetic group, cofactor. Classification of enzymes with two examples of each class.	(15)
Unit II	 LIPIDS: (05 L) Definition & classification with two examples of each class. Structure & functions of- a) Simple lipids- Fatty acids & triglycerides.	(15)

Reference Books:

- Lehninger, Principles of Biochemistry. 5th Edition (2008), David Nelson & Michael 1.
- Biochemistry Lubert Stryer, 5th Edition, W.H. Freeman and Company, New York. Fundamentals of Biochemistry Voet & Voet& Pratt. 2.
- 3.
- Fundamentals of Biochemistry J. L. Jain, S. Chand & Company Ltd, New Delhi. Biochemistry U. Satyanarayan, 3rd Edition, Books and allied (P) Ltd. 4.
- 5.
- Biochemistry S.C.Rastogi. 6.
- Text book of Biochemistry R. C. Dubey. 7.

SEMESTER-III

PAPER - II: BIOCHEMICAL TECHNIQUES

- Students learn about the basic techniques used for protein and DNA separation.
- Students get introduced to bioinformatics .
- Students learn about the principle and working of colorimeter and spectrophotometer.
- Students learn the chemical basis of various assay methods for biomolecules.
- Students learn the principles of frequently used clinical and research techniques
- Students get a closer view of practical biochemistry.

SEMESTER-III PAPER - II: BIOCHEMICAL TECHNIQUES

Total marks: 50 (40+10)		Credits: 2	Contact ho	urs: 30
Unit No.		Content		Lectures
	CHROMATO	SRAPHY (06 L)		
	Definition & c	classification, principle, technique & applications of		
	i)	Thin layer chromatography		
	ii)	Gel permeation chromatography		
	iii)	High pressure liquid chromatography		
	Selection of g	el, preparation of plate/column packing, application of sa	mple,	
	mechanism of	f separation, important applications & advantages of the n	nethods.	
	ELECTROPHO	DRESIS: (05 L)		
TT	Definition, electr	ophoretic mobility, factors affecting electrophoretic mobi	lity.	(15)
Unit I	Principle, technic	jue and applications of-		(15)
		1) Polyacrylamide gel electrophoresis: Native and SDS	3	
		2) Agarose gel electrophoresis		
	Propagation of ga	I plates application of sample machanism of sanaration	davaloning	
	Preparation of gel plates, application of sample, mechanism of separation, developing the plates, important applications and advantages of the methods		developing	
	BIOINFORMATICS (04 L)			
	Introduction to b	ioinformatics. Databases. Information sources (NCBI, GI	DB. and	
	MGD), Data retr	ieval tools (ENTREZ, OMIM and PubMed), Database sin	nilarity	
	searching (BLAS	T), Applications of Bioinformatics	5	
	ABSORPTION	SPECTROSCOPY (05 L)		
	Beer-Lambert's l	aw and its derivation, meaning of the terms- transmittanc	e,	
	absorbance, mola	ar and specific absorbance. Electromagnetic spectrum		
	Difference betwe	en photoelectric colorimeter and spectrophotometer		
	Advantages of sp	ectrophotometer over colorimeter.		
	ESTIMATION	OF BIOMOLECULES (05 L)		
	A) Carbohydrate	s - DNSA, anthrone, phenol-H ₂ SO ₄		
Unit II	B) Protein – BCA	A assay, Bradford assay, Lowery's assay.		
	C) Lipid – a	cid value, saponification value, ester value and iodi	ine number.	
	Liberman Burcha	ard method for cholesterol estimation.		
	D) Nucleic acids	– DPA method, orcinol method		
	MODERN TEC	HNIQUES (05 L)		
	A) Blotting te	chniques- Western, Southern & Northern blotting.		
	B) Polymeras	e chain reaction-technique & applications.		
	C) Enzyme li	nked immunosorbent assay (ELISA)-technique & applica	tions.	

Reference Books:

- 1. Protein purification Robert Scoopes
- 2. Instrumental Methods of Chemical Analysis Gurudeep R. Chatwal, Sham K. Anand (Himalaya Publishing House).
- 3. Handbook on Analytical Instruments -R. S. Khandpur. (McGraw Hill).
- 4. Biophysical Chemistry Upadhyay, Nath, Upadhyay (Himalaya Publishing House).
- 5. A Text Book of Biotechnology R. C. Dubey.
- 6. Practical Biochemistry Wilson & Walker.

SEMESTER-IV

PAPER - III: NUTRITION AND METABOLISM

- Students learn about the importance of nutrition, electrolyte and acid base balance.
- Students learn about important biochemical pathways of carbohydrate metabolism
- Students learn about mechanism and aspects related to oxidative phosphorylation.
- Students get introduced to mechanism of lipid metabolism
- Students get introduced to outline of nucleic acid metabolism.
- Students get introduced to general reactions of amino acid metabolism

SEMESTER-IV

PAPER - III: NUTRITION AND METABOLISM

Total m	narks: 50 (40+10) Credits: 2	Contact hours: 30
Unit No.	Content	Lectures
Unit I	 Nutrition and its components (05 L) A) Nutrition, balanced diet, calorific values of food and its measurement calorimeter) respiratory quotient, basal metabolic rate (BMR), Factors ef B) Electrolyte & acid base balance: 	t (bomb ffecting BMR. e in body (15) e of pyruvate, n dation of
Unit II	 palmitic acid and its energetics. Biological oxidation: (06 L) Bioenergetics-Exergonic and endergonic reactions, free energy, high energy compounds and their significance, ATP as a high energy compound. Mitochondrial respiration-components of respiratory chain, respiratory cion (Chacoupling hypothesis), inhibitors of electron transport chain. Metabolism of Nucleic acids (05 L) Sources of the atoms in the purine and pyrimidine molecules Outline of biosynthesis and degradation of purines and pyrimidines Metabolism of Amino acids (04 L) General reactions of amino acid metabolism viz. transamination, deamin decarboxylation. Urea cycle 	ergy chain, emiosmotic (15) nation,

Reference Books

- 1. Lehninger's Principles of Biochemistry –Nelson & Cox, 5th Edition, W.H. Freeman and Company, New York.
- 2. Fundamentals of Biochemistry Voet & Voet, 3rd Edition, W.H. Freeman and Company, New York.
- 3. Biochemistry Lubert Stryer, 5th Edition, W.H. Freeman and Company, New York.
- 4. Nutritional Biochemistry Dr.S.Ramkrishna & Dr. S. Vyankatrao.
- 5. Fundamentals of Biochemistry J. L. Jain, S. Chand & Company Ltd, New Delhi.
- 6. Biochemistry U. Satyanarayan, 3rd Edition, Books and allied (P) Ltd.

PAPER - IV:

MOLECULAR BIOCHEMISTRY & DISEASES

- Students learn mechanism of enzyme catalysis and various units of enzyme activity.
- Students get a closer view of enzyme kinetics.
- Students learn about basic tools of genetic engineering.
- Students understand the process of replication, transcription and translation in prokaryotes.
- They also learn about operon concept and its importance in gene regulation.
- Students get introduced to the subject of immunology.
- Students understand biochemical and immunological basis of: AIDS and diabetes mellitus.
- Students understand how the anti AIDS and hypoglycemic drugs work.

PAPER - IV: MOLECULAR BIOCHEMISTRY & DISEASES

Total marks: 50 (40+10)Credits: 2Contac	
Content	Lectures
 ENZYMOLOGY (06 L) A) Enzyme as a catalyst: concept of activation energy, Lock & key model, Induced fit hypothesis. Active site of enzyme and its features. B) Enzyme activity: Definition & Unit of enzyme activity, specific activity and turnover number, Factors affecting enzyme activity-pH, temperature and substrate concentration. C) Enzyme kinetics: Derivation of Michaelis-Menten equation for single substrate. Significance of Km and Vmax. Enzyme inhibition-irreversible, competitive and non competitive inhibition. 	
MOLECULAR BIOLOGY AND IMMUOLOGY (05 L) A) Molecular biology: Replication of DNA (semi conservative), transcription in prokaryotes. Translation in prokaryotes. Regulation of gene expression: constitutive & inducible genes. Operon concept, Lac operon in E. coli.	(15)
GENETIC ENGINEERING: (04 L) A) Genetic engineering: Restriction endonucleases, S1 nucleases, reverse transcriptase, cloning vectors-pBR322 and λ phage. Gene cloning technique illustrated with insulin gene cloning. Applications of generic engineering.	
 BASICS OF IMMUNOLOGY: (05) Natural & acquired immunity. Types and properties of antibodies, Structure of IgG. Phagocytosis and pinocytosis, Introduction to the cells involved in immune response: B cells and T cells BIOCHEMISTRY OF AIDS: (05 L) AIDS: Structure of HIV, transmission of HIV, immunological abnormalities in AIDS. Lysis of CD4 cells. natural course of AIDS- acute, chronic, crisis phases. Anti AIDS drugs-AZT, didanosine (structure & mechanism of action). BIOCHEMISTRY OF DIABETES MELLITUS (05 L) Structure of insulin, metabolic effects of insulin, mechanism of action of insulin. Types of diabetes mellitus, Symptoms, hyperglycemia, hypoglycemia, glucosourea and kidney threshold, Long term effects of diabetes: introduction to nephropathy, neuropathy, retinopathy 	(15)
	 Credits: 2 Contact h Contact h Content Content ENZYMOLOGY (06 L) A) Enzyme as a catalyst: concept of activation energy, Lock & key model, Induced fit hypothesis. Active site of enzyme and its features. B) Enzyme activity: Definition & Unit of enzyme activity, specific activity and turnover number, Factors affecting enzyme activity-pH, temperature and substrate concentration. C) Enzyme kinetics: Derivation of Michaelis-Menten equation for single substrate. Significance of Km and Vmax. Enzyme inhibition-irreversible, competitive and non competitive inhibition. MOLECULAR BIOLOGY AND IMMUOLOGY (05 L) A) Molecular biology: Replication of DNA (semi conservative), transcription in prokaryotes. Translation in prokaryotes. Regulation of gene expression: constitutive & inducible genes. Operon concept, Lac operon in E. coli. GENETIC ENGINEERING: (04 L) A) Genetic engineering: Restriction endonucleases, S1 nucleases, reverse transcriptase, cloning vectors-pBR322 and λ phage. Gene cloning technique illustrated with insulin gene cloning. Applications of generic engineering. BASICS OF IMMUNOLOGY: (05) Natural & acquired immunity. Types and properties of antibodies, Structure of IgG. Phagocytosis and pinocytosis, Introduction to the cells involved in immune response: B cells and T cells BIOCHEMISTRY OF ALDS: (05 L) AIDS: Lysis of CD4 cells. natural course of AIDS- acute, chronic, crisis phases. Ant AIDS drugs-AZT, didanosine (structure & mechanism of action of insulin. Types of diabetes mellitus, Symptoms, hyperglycemia, hypoglycemia, glucosourea and kidney threshold, Long term effects of diabetes: introduction to nephropathy, neuropathy, retinopathy and cardiovascular disease, Hypoglycemic drugs: metformin and sulfonylurea.

Reference Books

Cell and Molecular biology - P. K. Gupta.

- 1. Molecular Biology; R. Weaver; 2nd Edition, McGraw Hill.
- 2. Molecular Cell Biology; Lodish; 6th Edition; W. H. Freeman & Company.
- 3. Gene VII; Benjamin Lewin; Pearson Education.
- 4. Elements of Biotechnology P. K. Gupta.
- 5. Genetic engineering Sandhya Mitra.
- 6. Biotechnology B. D. Singh.
- 7. Immunology .- Kuby.
- 8. Essential Immunology- Roitt
- 9. Bioinformatics : Principle and applications Harshawardhan P. Bal.
- 10. Introduction to Bioinformatics T. K. Attwood & D. J. Parry- Smith

B.Sc. – II Biochemistry

Practical Course outcomes

- Students learn the principle and technique of separation by chromatography.
- Students learn the basics of isolation and characterization of biological samples.
- Students understand the application of Beer Lamberts law in colorimetric estimation technique.
- Students understand the principle of estimation of biomolecules by colorimetry.
- Students learn the basics of titration and volumetric estimation of different components.
- Students learn the basics of qualitative estimation of biomolecules using specific tests.
- Students get introduced to an enzyme assay, its quantification and factors affecting on it.

B.Sc. – II Biochemistry

Practical Course

Note: 1. Use of Electronic/single pan/digital balance is allowed.

- 2. Use of scientific calculator is allowed.
- 3. Use SI units whenever possible.

Part I

- 1. a) Fundamentals of Biochemical analysis.
 - b) Control and accuracy.

2. Separation/characterization (Any Two):

- a) Paper chromatographic separation and identification of amino acids from binary mixture.
- b) Paper chromatographic separation and identification of carbohydrates from binary mixture.
- c) Thin layer chromatographic separation and identification of amino acids/carbohydrates from binary mixture.

3. Preparation of biological samples (Any Two):

- a) Isolation and characterization of starch from potatoes
- b) Isolation and characterization of casein from milk
- c) Isolation and characterization of albumin and globulin from egg yolk.

4. Colorimetric estimations (Any five):

- a) Estimation of protein by Biuret method and verification of Beer-Lamberts law.
- b) Estimation of inorganic phosphate in blood by Fiske-Subbarao method.
- c) Estimation of reducing sugar by DNSA method.
- d) Estimation of RNA by Bial's Orcinol method.
- e) Estimation of creatinine in urine.
- f) Estimation of blood urea by DAM method.
- g) Estimstion of DNA by Diphynylamine method.

Part II

1. Demonstration Experiments:

- a) Soxhlet extraction of lipids from ground nuts/ egg yolk.
- b) Separation of indicators/ serum proteins by paper/ cellulose acetate electrophoresis.
- c) Antigen antibody interaction by Ouchterlony immunodiffusion procedure.
- d) Study of 3D structure and function relation of an Enzyme (Using Bioinformatics Tool).

2. Volumetric estimations (Any Four):

- a) Estimation of glycine by formal titration.
- b) Estimation of lactose in milk by Fehlings methods.
- c) Estimation of vitamin C in biological samples and tablets by 2,6dichlorophenol indophenols method.
- d) Estimation of saponification value of oil.
- e) Estimation of iodine number of oil.
- f) Estimation of acid value of oil.

3. Qualitative analysis:

- a) Carbohydrates xylose, glucose, fructose, lactose, maltose, sucrose, starch.
- b) Lipids oleic acid, palmitic acid, cholesterol, glycerol.
- c) Detection of any Three enzymes by qualitative tests (Urease, invertase, amylase, phenol oxidase, alkaline phosphatase, catalase)

d) Abnormal constituents in urine (blood, reducing sugar, proteins, bilirubin, ketone bodies).

4. Study of Enzymes

- a) Immobilization of baker's yeast cells by gel entrapment for invertase activity.
- b) Quantitative Estimation of amylase activity.
- c) Effect temperature/pH/substrate concentration on amylase enzyme

List of reference books for practical

- 1. Laboratory manual in Biochemistry J. Jaynaraman
- 2. Practical Biochemistry David Plummer
- 3. Hawk's physiological chemistry Oser
- 4. A manual of laboratory technique (Ed) N. Raghuramulu, K. Madhavan Nair & S. Kalyansundaram
- 5. Biochemistry methods Sadasivan & Manikam.
- 6. Introductory Practical Biochemistry Sawhney S. K. and Ranabir Singh
- 7. Viva and Practical Biochemistry A. C. Deb.

B.Sc. Part-II

PRACTICAL EXAMINATION IN BIOCHEMISTRY

The practical examination of B.Sc. Part II in Biochemistry will be of 12 hours duration and will be conducted in two successive days, 6 hours per day.

The total practical examination will be of 160 Marks. The distribution of marks will be as follows.

1. Colorimetric estimation 20 2. Isolation of biological samples 20 3. Paper chromatography /Separation method 20 4. Volumetric estimation 20 5. Qualitative analysis of 20 a) Carbohydrate b) Lipid/enzyme c) Abnormal Urine components 6 Study of Enzyme 20 a)immobilization b) Activity Quantification c) Effect of pH/temperature 7 Oral 10 8 Journal 20 9 Visit Report of Study tour 10

The practical batch will be of maximum 24 candidates. The batch will be divided in two groups A & B. Group A will consist of 12 candidates while group B will have 12 candidates.

If the number of candidates in batch is less than 20 the number should be divided in two equal groups A & B. Any number remaining will be placed in group B.

Practical examination timing:

11.00 am to 2.00 pm 2.30 pm to 5.30 pm

Recess 2.00 pm to 2.30 pm

Nature of Question Paper for choice based credit system (CBCS) Semester Pattern

Faculty of Science (w. e. f. June2023)

TotalMarks-40

Time: -2hrs.

Instructions:

- All questions are compulsory.
 Draw neat diagrams and give equations wherever necessary.
 Figures to the right indicate full marks.
- 4. Use of logarithmic table and calculator is allowed.

Q. No.1) Multiple choice questions	(08)
a) b) c) d) 2) 3) 4) 5) 6) 7) 8)	
Q.No.2) Answer any four of the following	(08)
i)	
ii)	
111) iv)	
v)	
vi)	
Q.No.3 Attempt any Two of the following	(08)
i) ii) iii)	
Q. No.4) Answer any Two of the following	(08)
ii) iii)	
Q.No.5) Answer any one of the following	(08)
11)	